Abstracts of papers presented at the 56th Fujihara Seminar as

International Conference on Molecular Mechanism of Intracellular Transport: The Roles of Kinesin and Dynein Superfamily Proteins

The Molecular Motor Conference 2007

August 23-26, 2007

At Grand Hotel New Oji in Tomakomai, Hokkaido, Japan

Organizer Nobutaka Hirokawa

The Fujihara Foundation of Science

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56th Fujihara Seminar The Molecular Motor Conference 2007

Thursday, August 23 - Sunday, August 26, 2007

Thursday, 23 rd	7:00 pm	Registration, Poster hanging	
	7:30 pm	WELCOME PARTY in Foyer (2 nd Floor)	
Friday, 24 th	8:00 am	Opening Remarks	
	8:30 am	1 Intracellular transport (1-4)	
	1:00 pm	POSTER SESSION I (A-N; 21-38)	
	2:00 pm	РНОТО	
	2:30 pm	2 Motility (5-8)	
	6:30 pm	RECEPTION at Harunire (1st Floor)	
Saturday, 25 th	8:30 am	3 Mitosis (9-12)	
	1:00 pm	POSTER SESSION II (O-Y; 39-55)	
	2:00 pm	EXCURSION	
Sunday, 26 th	8:30 am	4 Transport (13-16)	
	1:00 pm	5 Cilia and Dynein (17-20)	
	4:30 pm	6 Dynein (Short Talks)	
	6:30 pm	Closing Remark	
	7:00 pm	BANQUET at Grandview (16th Floor)	

All sessions are located at Fuyou Hall (2nd Floor).

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FRIDAY, August 24

8:00 am – 8:	30 am Opening Remarks	
8:00 am	Greeting SOJI BAN, Executive Director, The Fujihara Foundation of Science	
8:10 am	Introductory Remark SUSUMU NISHIMURA, Visiting Professor, Center for TARA, Univ. of Tsukuba	
8:20 am	Introductory Remark NOBUTAKA HIROKAWA, Graduate School of Medicine, University of Tokyo	
8:30 am – No	SESSION 1 Intracellular transport	
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2:00 pm

РНОТО

2:30 pm - 6:00 pm

SESSION 2 Motility

- Chairpersons: Sharyn A. Endow Robert Cross
- 2:30 pm ROBERT CROSS

Alonso M¹), Yajima J²), Carter N¹) & Cross R¹) ¹/Marie Curie Research Institute, ²/Department of Physics, Gakashuin University Mechanics of Kinesin-1

3:15 pm TOSHIO YANAGIDA

Komori Y^{1,2)}, Iwane AH^{1,3)} & Yanagida T^{1,2,3)} ¹⁾Laboratories for Nanobiology, Graduate School of Frontier Biosciences, Osaka University, ²⁾Formation of Soft Nano-Machines, CREST JST, ³⁾Department of Physiology and Biosignaling, Graduate School of Medicine, Osaka University **Myosin-V Makes Two Brownian 90° Rotations per 36 nm-Step**

4:00 pm COFFEE

4:30 pm Sharyn A Endow

Endow SA¹⁾, Zou J¹⁾, Yankel CD¹⁾, Amos LA²⁾ & Hirose K³⁾ ¹⁾Duke University Medical Center, ²⁾MRC Laboratory of Molecular Biology, ³⁾National Institute of Advanced Industrial Science and Technology (AIST) Kinesin Motor Structural Changes and Function in Meiosis

5:15 pm JONATHON HOWARD Howard J, Brouhard G, Helenius J, Hyman AA, Stear J & Varga V Max Planck Institute of Molecular Cell Biology and Genetics Regulation of Microtubule Length by Depolymerizing Kinesins and Polymerases

SATURDAY, August 25

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2:00 pm - 6:30 pm

EXCURSION

SUNDAY, August 26

8:30 am – No	on SESSION 4 Transport	
Chairpersons	: Vladimir I Gelfand Gary Banker	
8:30 am	GARY BANKER Banker G, Huang CF, Petersen J & Kaech S Center for Research on Occupational and Environmental Toxicology, Oregon Health & Science University The Long-Range Carriers that Transport Polarized Membrane Proteins to Axons and Dendrites	13
9:15 am	KRISTEN VERHEY Cai D ¹ , Reed N ¹ , Meyhofer E ² & Verhey K ¹ Departments of ¹ Cell and Developmental Biology and ² Mechanical Engineering, University of Michigan, USA Regulation of Kinesin-1 Transport by Microtubule Post-Translational Modifications	14
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1:45 pm	JOEL ROSENBAUM Rosenbaum J Dept. Molecular, Cellular and Developmental Biology, Yale University Intraflagellar Transport Protein, IFT27, a Small G-Protein, Controls Both the IFT-Dependent Assembly of the Cilium, and Cell Division	18
2:30 pm	RICHARD VALLEE Tsai JW, Seale GE & Vallee RB Columbia University, College of Physicians and Surgeons, Dept. of Pathology and Cell Biology Multiple Roles for Cytoplasmic Dynein and LIS1 in Neuronal Migration and Axonal Pathfinding	19
3:15 pm	RITSU KAMIYA Yagi T, Kikushima K & Kamiya R Department of Biological Sciences, Graduate School of Science, University of Tokyo Novel Species and Novel Motor Properties of Axonemal Dyneins	20
4:00 pm	COFFEE	
4:30 pm – 6:	30 pm SESSION 6 Dynein (Short Talks)	
Chairpersons	s: Kazuhiro Oiwa Ritsu Kamiya	
4:30 pm	Kamimura S ¹⁾ & Iwamoto H ²⁾ ¹⁾ Department of Life Sciences, Graduate School of Arts and Sciences, University of Tokyo & ²⁾ Life and Environmental Division, SPring-8, JASRI	

SESSION 5 Cilia and Dynein

1:00 pm – 4:00 pm

Dynein Arm Arrangement in Flagellar Axonemes and Its Dynamic Change Revealed by Small-Angle X-ray Diffraction Analysis

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	Department of Life Sciences, Graduate School of Arts and Sciences, University of Tokyo	
	Dissection of Intra-Molecular Communications Between the Catalytic Head and Microtubule-Binding Site in the Dynein Heavy Chain	31
5:10 pm	Nguyen H ¹), Kaya M ¹), Kon T ²), Sutoh K ²) & Higuchi H ¹)	
	¹⁾ Biomedical Engineering Research Organization, Tohoku University, ²⁾ Department of Life Sciences, Graduate School of Arts and Sciences, University of Tokyo	
	The Contribution of Individual Dynein Head to the Processive Movement Generated by Two Single Headed Dynein	36
5:30 pm	Oiwa K	
	Kobe Advanced ICT Research Center, National Institute of Information and Communications Technology; Graduate School of Life Science, University of Hyogo	
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	Department of Biological Sciences, Graduate School of Science, University of Tokyo	
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6:10 pm	Ueno H ¹⁾ , Yasunaga T ²⁾ , Shingyoji C ³⁾ & Hirose K ¹⁾	
	¹⁾ Research Institute for Cell Engineering, AIST, ²⁾ Department of Biochemical Engineering and Science, Kyushu Institute of Technology, ³⁾ Department of Biological Sciences, Graduate School of Science, University of Tokyo	
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6:30 pm

Closing Remark

7:00 pm

BANQUET

NOBUTAKA HIROKAWA

Integrative Biology of Kinesin Superfamily Molecular Motors, KIF 1A, KIF 3 and KIF 4: Structure, Dynamics and Functions

Hirokawa N

Department of Cell Biology and Anatomy, Graduate School of Medicine, University of Tokyo, Hongo 7-3-1, Bunkyo-ku, Tokyo, 113-0033 Japan

The intracellular transport is essential for cell morphogenesis, functioning and survival. To elucidate this mechanism we have identified and been characterizing kinesin superfamily proteins, KIFs, using multidisciplinary and integrative approaches combining molecular cell biology, molecular genetics, biophysics, X-ray crystallography and cryoelectron microscopy. In this seminar I will focus on KIF 3, KIF 4 and KIF 1A.

To know the in vivo function of KIF3 in brain we conditionally knocked out KAP3, an associated protein of KIF3 to suppress function of KIF3 in brain. In mutant brain progenitor cells proliferate and invade upper layers to cause tumor like rossete formation. In the KAP 3 knock out cells transport of N-cadherin and beta-catenin from Golgi to plasma membrane was affected. We found that the cargoes of KIF 3 contain N-cadherin and beta catenin and that KIF 3 controls a balance between beta catenin in the cytoplasm and it in the plasma membrane working as cell-cell adhesion molecule. Beta catenin in the cytoplasm tends to get in the nucleus to work as a transcriptional factor together with T cell factor and enhances cell proliferation. Thus this study suggests a potential tumor-suppressing activity of KIF 3.

In brain development, apoptosis is a physiological process that controls the final numbers of neurons. We found that the activity dependent prevention of apoptosis in juvenile neurons is regulated by KIF 4. The C-terminal domain of KIF 4 is a module that suppresses the activity of poly(ADP-ribose)polymerase-1(PARP-1), a nuclear enzyme. When neurons are stimulated by membrane depolarization, calcium signaling mediated by CaMKII induces dissociation of KIF 4 from PARP-1, resulting in upregulation of PARP-1 activity, which supports neuron survival. After dissociation from PARP-1, KIF 4 enters into the cytoplasm from the nucleus and moves to the distal part of neuritis in a microtubule-dependent manner. We suggest that KIF 4 controls the activity-dependent survival of postmitotic neurons by regulating PARP-1 activity in brain development.

In terms of the mechanism of motility the slow rate-limiting ADP release from free kinesin is accelerated by more than 10⁴-fold following interaction with MT. This MT-dependent regulation serves as a chemical checkpoint to suppress futile consumption of ATP by free kinesin. Thus, ADP release is a key fundamental regulatory step for kinesin. We propose an atomic mechanism for this regulation based on five new crystal structures of ADP-releasing intermediates of the monomeric kinesin KIF1A and supporting kinetic measurements of mutant KIF1As.

VIKI ALLAN

Kinesin-1 and Endoplasmic Reticulum Motility

Allan V & Wozniak M

Faculty of Life Sciences, University of Manchester, The Michael Smith Building, Oxford Road, Manchester, M13 9PT, United Kingdom

Kinesin-1 is thought to drive the movement of many different cargoes, including membrane-bound organelles such as the endoplasmic reticulum (ER). Kinesin-1-driven movement towards microtubule plus ends would transport the ER tubules towards the cell periphery in most cells, and this movement, together with the fusion of ER tubules with each other, is proposed to generate the typical reticular-tubular membrane network. Since kinesin-1 transports multiple cargoes, an important question is how the motor associates with these different structures. One possibility is that identical motor molecules bind to distinct cargo-specific receptors, such as kinesin heavy chain (KHC) binding to kinectin in the ER. However, kinesin light chains (KLCs) also bind to certain cargo molecules, and KLC1 exists in at least 8 splice isoforms, which differ only at their C-termini. We have tested whether distinct KLC1 isoforms play a role in targeting kinesin-1 to different organelles. Using a combination of in vitro motility assays and biochemical approaches, we have found that KLC1B is involved in targeting and function of kinesin-1 on the ER, whereas KLC1D is required for the movement of vesicles in a Golgi membrane fraction. We have confirmed these results using a new assay for ER motility that uses semiintact cells. However, when transient transfection is used to express KHC or KLC fragments in cells, or when KHC expression is knocked down using RNAi, only subtle effects are seen on ER morphology and motility. This may be because ER tubules can extend in mulliple ways-via kinesin or dynein, or by attaching to growing microtubules. Moreover, it is clear that other motile organelles such as endosomes, lysosomes, mitochondria and peroxisomes, may form transient interactions with the ER, and therefore extend ER tubules indirectly using motors other than kinesin-1.

The 56th Fujihara Seminar Grand Hotel New Oji in Tomakomai. August 23-26, 2007

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